Plant Gene Register

Chloroplastic Glutamine Synthetase from Brassica napus

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In higher plants, the GS/glutamate synthase cycle provides the only efficient pathway for the conversion of inorganic nitrogen to the organic form (Stewart et al., 1980). In this connection, GS catalyzes the ATP-dependent condensation of ammonia with glutamate to yield Gln. Ammonia, the ultimate inorganic form of nitrogen available to the plant, is released by a variety of physiological processes in different organs and subcellular locations within the plant (Miflin and Lea, 1980). Photorespiration in the leaves of C₃ plants represents an important secondary source of ammonia that is reassimilated by GS activity (GS₂) located in the stroma of chloroplasts (Woo et al., 1982; Wallsgrove et al., 1987).

To study the plastidic GS isoenzyme of *Brassica napus*, we screened a λ -ZAPII cDNA library constructed using poly(A)⁺ mRNA obtained from rape leaves (Table I). The pGS185 cDNA clone from pea (Tingey et al., 1988) was used as a hybridization probe to identify positive phage plaques. Partial sequence analysis and restriction mapping of isolated clones indicated that the coding regions of all cDNAs were identical. The sequence of the longest cDNA, a putative full-length clone of 1575 bp, has been called BnGS18. The nucleotide sequence of this cDNA shows the occurrence of an open reading frame of 1284 nucleotides, a 94-nucleotide 5' untranslated region, and a 177-nucleotide noncoding 3' end followed by a polyadenylation stretch of 20 nucleotides.

The open reading frame encodes a precursor protein for a GS₂ subunit of 428 amino acids. If we assume the same processing site for the removal of the transit peptide, as identified by sequence similarity with other plastidic GS enzymes (Lightfoot et al., 1988; Tingey et al., 1988; Peterman and Goodman, 1991; Becker et al., 1992), the molecular mass of the mature protein would be nearly 42 kD. This is in agreement with the apparent molecular mass of the GS2 subunit as determined by western analysis of rape leaf soluble proteins (data not shown). The amino acid sequence of the mature GS₂ subunit from B. napus exhibits 94% identity with the GS₂ sequence of Arabidopsis thaliana, another species of Brassicaceae (Peterman and Goodman, 1991). Furthermore, comparison with legume species revealed 88 and 87% similarity with the bean (Lightfoot et al., 1988) and pea sequence (Tingey et al., 1988), respectively.

A comparison of the transit peptide sequences shows less conservation, although there are regions of similarity. The first 147 nucleotides of the open reading frame translate into a polypeptide with a high proportion of basic amino acid residues (18%) that is completely devoid of acidic amino

Organism

Brassica napus var Arabella.

Gene Product:

Chloroplastic GS (EC 6.3.1.2).

Techniques:

Screening of λ -ZAPII cDNA library constructed from green leaves using a pea cDNA probe specific for the plastidic isoenzyme (clone pGS 185); double-stranded plasmid (pBluescript) dideoxynucleotide sequencing of both strands by primer jumping.

Method of Identification:

Comparison of the nucleotide sequence and the deduced amino acid sequence with previously described GS sequences from other higher plants.

Features of cDNA:

The clone BnGS18 is 1575 nucleotides in length and consists of a 94-nucleotide 5' untranslated region, a 1284-nucleotide open reading frame, and a 197-nucleotide 3' untranslated segment.

Regulation:

Light-stimulated accumulation in green tissue of a 1.7-kb transcript as determined by northern analysis.

Features of Protein:

This cDNA contains an open reading frame of 428 amino acids (M, 47,342), which encodes a 49-amino transit peptide $(M_r 5,382)$ and a 379-amino acid mature protein $(M_r 41,978)$.

Antibodies:

Polyclonal antibodies raised against GS₂ from Sinapis alba crossreact with GS₂ from Brassica napus.

Subcellular Location:

Chloroplastic.

acids; both are characteristics of chloroplast transit peptide sequences (Karlin-Neumann et al., 1986). Further experiments are currently in progress to isolate additional GS-specific transcripts from a root cDNA library of *B. napus*. When this work is complete it will be possible to examine the regulation and expression of GS gene(s) from a nonlegume species.

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Table I. Characteristics of GS cDNA from Brassica napus

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Abbreviations: GS, glutamine synthetase; GS₂, chloroplastic GS.

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LITERATURE CITED

- Becker TW, Caboche M, Carrayol E, Hirel B (1992) Nucleotide sequence of a tobacco cDNA encoding plastidic glutamine synthetase and light inducibility, organ specificity and diurnal rhythmicity in the expression of the corresponding genes of tobacco and tomato. Plant Mol Biol 19: 367–379
- Karlin-Neumann GA, Tobin EM (1986) Transit peptides of nuclearencoded chloroplast proteins share a common amino acid framework. EMBO J 5: 9–13
- Lightfoot DA, Green NK, Cullimore JV (1988) The chloroplast-located glutamine synthetase of *Phaseolus vulgaris* L.: nucleotide sequence, expression in different organs and uptake into isolated chloroplasts. Plant Mol Biol 11: 191–202
- Miflin BJ, Lea PJ (1980) Ammonia assimilation. In BJ Miflin, ed, The

- Biochemistry of Plants, Vol 5. Academic Press, San Diego, CA, pp 169-202
- Peterman TK, Goodman HM (1991) The glutamine synthetase gene family of *Arabidopsis thaliana*: light-regulation and differential expression in leaves, roots and seeds. Mol Gen Genet 230: 145–154
- Stewart GR, Mann AF, Fentem PA (1980) Enzymes of glutamate formation: glutamate dehydrogenase, glutamine synthetase, and glutamate synthase. *In BJ Miflin*, ed, The Biochemistry of Plants, Vol 5. Academic Press, San Diego, CA, pp 271–327
- Tingey SV, Tsai FJ, Edwards JW, Walker EL, Coruzzi GM (1988) Chloroplast and cytosolic glutamine synthetase are encoded by homologous nuclear genes which are differentially expressed in vivo. J Biol Chem 263: 9651–9657
- Wallsgrove RM, Turner JC, Hall NP, Kendall AC, Bright SWJ (1987) Barley mutants lacking chloroplast glutamine synthetase—biochemical and genetic analysis. Plant Physiol 83: 155–158
- Woo KC, Morot-Gaudry JF, Summons RÉ, Osmond CB (1982) Evidence for the glutamine synthetase/glutamate synthase pathway during the photorespiratory nitrogen cycle in spinach leaves. Plant Physiol 70: 1514–1517